EXPERIMENT 7 – GENE EXPRESSION STUDY IN *ARABIDOPSIS THALIANA* (GENE TWO)

Purpose: To determine mRNA accumulation patterns of genes encoding transcription factors in *Arabidopsis* leaves and siliques.

STRATEGY

Use the cDNA solutions that you generated in the Experiment 3 with the RT-PCR primers for gene TWO.

PROCEDURE

CARRYING OUT PCR AMPLIFICATION (RT-PCR) ANALYSIS

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Purpose: To amplify DNA template corresponding to mRNA that is either absent or present at different levels in different plant organs throughout plant development.

Materials and Reagents Needed:

- Reverse transcription (+RT & -RT) reactions
- 10X Ex-Taq buffer (Takara Mirus Bio)
- dNTP mix (Takara Mirus Bio)
- **12 μM** Gene-specific **RT-PCR Fw** primers
- **12 μM** Gene-specific **RT-PCR Rv** primers
- **12 μM Tubulin Fw** primers
- **12 μM Tubulin Rv** primers
- Sterile water
- Ex-Taq DNA polymerase (Takara Mirus Bio, 5 U/ μL)
- Agarose
- 1X TAE buffer
- 100-bp DNA ladder (Invitrogen)
- 6X Loading buffer containing ONLY **Xylene Cyanol**
- 0.2 mL PCR tubes
- 1.5 mL microcentrifuge tubes
- Microcentrifuge tube racks
- PCR Machine (Bio-Rad MyCycler or PE 9700)
- Gel apparatus
- Power supplies

PROCEDURE

16. Get ice from the icemaker in room 2911 or 3906.
17. Determine **how many RT reactions**, including **+RT's** and **-RT's**, will be amplified.
18. Make a **Table** with information sush as **tube #**, **plant organ(s)**, and **+RT's/-RT's** (see the **example** Table below)

Tube #	**1**	**2**	**3**	**4**	**5 (Positive)**	**6 (Negative)**
Organ & RT	Leaf +RT	Leaf -RT	Silique +RT	Silique -RT	Genomic DNA	Sterile Water

19. Label on the **lids** and **sides of SIX 0.2 mL PCR tubes** with **Number** and **your initial**.
20. Put the labeled tubes on a PCR rack sitting **on ice**.
21. Prepare a **master mix** in a **1.5-mL microcentrifuge tube** for the **number** of **PCR solutions** being carried out **plus** 1 **extra solution volume** as followings: **(How many reactions are carried out?)**

	Mmix for ONE Reaction	**Mmix** for **7** Reactions
10X Ex-Taq buffer	5.0 µL	35.0 µL
dNTP mix	4.0 µL	28.0 µL
12 µM RT-PCR Gene-specific **Fw** primer	1.0 µL	7.0 µL
12 µM RT-PCR Gene-specific **Rv** primer	1.0 µL	7.0 µL
***12 µM Control** (**Tubulin**) **Fw** primer	1.0 µL	7.0 µL
***12 µM Control** (**Tubulin**) **Rv** primer	1.0 µL	7.0 µL
Sterile water	34.6 µL	242.2 µL
Ex-Taq DNA Polymerase (5 U/µL)	0.4 µL	2.8 µL
Total Volume	**48.0** µL	**336.0 µL**

* ***Control primers*** *are used to ensure that the absence of Gene-specific PCR product in* ***the +RT samples*** *is NOT due to technical mistakes.*

22. Pipet **48 µL** of the **master mix** to the labeled tubes and **2 µL** of **appropriate RT** to each of the tubes shown on the table below. Mix the contents by pipetting **gently** up and down for five times.

Tube #	**1**	**2**	**3**	**4**	**5 (Positive)**	**6 (Negative)**
Mmix	**48 µL**	**48 µL**	**48 µL**	**48 µL**	**48 µL**	**48 µL**
Leaf +RT	**2 µL**	0 µL	0 µL	0 µL	0 µL	0 µL
Leaf -RT	0 µL	**2 µL**	0 µL	0 µL	0 µL	0 µL
Silique +RT	0 µL	0 µL	**2 µL**	0 µL	0 µL	0 µL
Silique -RT	0 µL	0 µL	0 µL	**2 µL**	0 µL	0 µL
0.2 ng/µL Genomic DNA	0 µL	0 µL	0 µL	0 µL	**2 µL**	0 µL
Water	0 µL	0 µL	0 µL	0 µL	0 µL	**2 µL**
Total Volume	**50 µL**	**50 µL**	**50 µL**	**50 µL**	**50 µL**	**50 µL**

23. Carry out PCR reactions with the **RT-PCR program** containing the following profile: 1 cycle of 96°C, 3 min. → 40 cycles of 94°C, 10 sec./60°C, 30 sec./72°C, 45 sec. → 1 cycle of 72°C, 4 min. → 4°C, ∞.
24. Prepare **100 mL** of **1.5% agarose** gel in **1X TAE** buffer as usual (Use a **20-tooth** comb).

    <u>Note:</u> *The **percentage** of **agarose gels** depends on the **difference** in **size** of **two PCR products**. If there is **at least 100-bp difference** between two PCR products, then use a **1% agarose gel**. However, if there is **50-100 bp difference** between two PCR products, then use **1.5-2% agarose gel**. For example, the size of PCR products is 0.6 kb and 0.55 kb for the control and gene A, respectively. The **2.0%** agarose gel resolves these two PCR products as **two** discreet DNA bands whereas the **1.0%** agarose gel shows these two PCR products as a **single** DNA band.*
25. Label 1.5 mL microcentrifuge tubes according to the PCR solutions being performed.

26. Add to the labeled 1.5 mL microcentrifuge tubes **20 µL** of **PCR solution** and **3 µL** of **6X loading dye containing ONLY xylene cyanol**. *Note: 20 µL of PCR solution was loaded so that you can see the PCR products clearly.*
27. Load samples on a **1.5%** agarose gel along with **10 µL** of **50 ng/µL 100-bp ladder** solution. Record RNA loading pattern.

—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
1	**2**	**3**	**4**	**5**	**6**	**7**	**8**	**9**	**10**	**11**	**12**	**13**	**14**	**15**	**16**	**17**	**18**	**19**	**20**

28. Run the gel at **105 volts** for **1-2 hours** or until the front dye (bromophenol blue) is about two-thirds of the gel.

    Starting time:

    Ending time:
29. Take a picture of the gel.
30. Analyze the data.

    How many DNA fragments do you see on the gel?

    Is there different brightness between the fragments from one organ to the other?

    What are the sizes of DNA fragments?

    What is the size of the PCR fragment corresponding to mRNA of the gene of interest?

    What is a conclusion on gene expression of the gene of interest for the tested plant organs, leaves and Siliques?

    Are the RT-PCR results correlated to the GeneChip data?