

**Supplemental Figure 1: SUVR1, SUVR2 and/or SUVR4 are required for nucleolar dominance in *A. suecica***

(A) Phylogenetic relationships among SUVR proteins, displayed as a Neighbor-Joining tree with bootstrap values.

(B) RNA was extracted from wild-type plants or T1 transformants expressing amiRNAs targeting *SUVR1/2/4* (B, left panel) or *SUVR1* alone (B, right panel). *A. thaliana*-derived rRNA gene transcripts were detected by RT-CAPS. Transgenic *A. suecica* expressing double-stranded RNA that targets *DRM2* and disrupts nucleolar dominance was used as a positive control. After RT-PCR amplification of the ITS1 region, amplicons were subjected to *HhaI* digestion to distinguish *A. thaliana*-derived from *A. arenosa*-derived rRNA genes. Reactions in which reverse transcriptase was omitted serve as negative controls (-RT).

(C) Specificity of *SUVR4* amiRNA targeting. To test for off-target effects, RT-PCR was conducted using primers specific for *SUVH4*, *SUVH5*, *SUVH6*, *SUVR1*, *SUVR2* or *ACT2* mRNAs. RNA was purified from wild-type (1/8x, 1/2x or 1x amounts) and four *SUVR4* amiRNA lines (1x RNA amounts).

**Supplemental Figure 2: SUVH4, SUVH5 and SUVH6 are not implicated in nucleolar dominance in *A. suecica*.**

A) RT-PCR analysis of *SUVH4* mRNA levels in T2 progeny of two independent *SUVH4* amiRNA lines, compared to wild-type (WT). Actin (*ACT2*) RT-PCR reactions control for the amount of RNA analysed.

- B) RT-PCR analysis of *SUVH5* and *SUVH6* mRNA levels in eight transgenic plants expressing an amiRNA that targets both *SUVH5* and *SUVH6*, compared to wild-type (WT) and actin controls.
- C) RT-CAPS analysis of *A. arenosa* and/or *A. thaliana*-derived rRNA gene transcripts in *A. suecica* wild-type or *SUVH4* knockdown lines. The derepression of *A. thaliana*-derived rRNA genes (arrow) in a *DRM2* RNAi line serves as a positive control for loss of nucleolar dominance. Actin (*ACT2*) RT-PCR reactions control for the amount of RNA analysed.
- D) RT-CAPS analysis of *A. arenosa* and/or *A. thaliana*-derived rRNA gene transcripts in *A. suecica* wild-type or transgenic plants expressing an amiRNA targeting both *SUVH5* and *SUVH6*. A *DRM2* RNAi line serves as a positive control for loss of nucleolar dominance. Actin (*ACT2*) RT-PCR reactions control for the amount of RNA analysed.
- E) RT-CAPS analysis of *A. arenosa* and/or *A. thaliana*-derived rRNA gene transcripts in *A. suecica* wild-type or *SUVH5* or *SUVH6* knockdown lines. A *DRM2* RNAi line serves as a positive control for loss of nucleolar dominance. Actin (*ACT2*) RT-PCR reactions control for the amount of RNA analysed.

**Supplemental Figure 3: Quantitative PCR analyses of *A. thaliana*-derived rRNA genes in wild-type and *SUVR4*-amiRNA plants.**

Specific TaqMan probes were used to evaluate the relative amounts of *A. thaliana* versus *A. arenosa*-derived rRNA gene transcripts by quantitative RT-PCR in wild-type plants or or plants expressing an amiRNA targeting *SUVR4* mRNAs.

**Supplemental Figure 4: H3K27me1 association with rRNA genes**

Data derived from a genome-wide ChIP-sequencing study of H3K27me1- associated regions (Jacob et al. 2010) shows the distribution of H3K27me1 throughout an rRNA gene repeat in wild-type *A. thaliana*, ecotype Col-0.

## Supplemental Materials and Methods:

### Primers for amiRNA production

IIFPamiratx12	GATTATAATCATCGGTCAGCGCCTCTCTCTTTTGTATTCC
IIIFPamiratx12	GAGGCGCTGAACTATGATTATAATCAAAGAGAATCAATGA
IIIFPamiratx12	GAGACGCTGACCTAAGATTATTATCACAGGTCGTGATATG
IVFPamiratx12	GATAATAATCTTAGGTCAGCGTCTCTACATATATATTCCT
IIFPamiratx345	GATCAAGCACATGTCACGTGAACTCTCTCTTTTGTATTCC
IIIFPamiratx345	GAGTTCACGTGACATGTGCTTGATCAAAGAGAATCAATGA
IIIFPamiratx345	GAGTCCACGTGACATCTGCTTGTTTACAGGTCGTGATATG
IVFPamiratx345	GAACAAGCAGATGTCACGTGGACTCTACATATATATTCCT
IImiRSUVH1	GATTCTAAATGACCTTAAAGGGGTCTCTCTTTTGTATTCC
IIImiRSUVH1	GACCCCTTTAAGGTCATTTAGAATCAAAGAGAATCAATGA
IIImiRSUVH1	GACCACTTTAAGGCTTTTATAGATTACAGGTCGTGATATG
IVmiRSUVH1	GAATCTAAAAGACCTTAAAGTGGTCTACATATATATTCCT
IIFPamIRSUVH2	GATATATGCAAACCTCACGGTCTCTCTTTTGTATTCC
IIIFPamIRSUVH2	GAACCGTGGTGAGTTTGCATATATCAAAGAGAATCAATG
IIIFPamIRSUVH2	GAACAGTGGTGAGTTAGCATATTTTACAGGTCGTGATAT
IVFPamIRSUVH2	GAAATATGCTAACTCACCCTGTTCTACATATATATTCCT
IImiRSUVH3	GATAATTATATAGTCAATGCCGGTCTCTCTTTTGTATTCC
IIImiRSUVH3	GACCGGCATTGACTATATAATTATCAAAGAGAATCAATGA
IIImiRSUVH3	GACCAGCATTGACTAAATAATTTTACAGGTCGTGATATG
IVmiRSUVH3	GAAAATTATTTAGTCAATGCTGGTCTACATATATATTCCT
IIFPamIRSUVH1&3	GATACATATAAAAAGAACCGGCACTCTCCTCTTTGTATTCC
IIIFpamIRSUVH1&3	GAGTGCCGGTCTTTTATATGTATCAAAGAGAATCAATGATGA
IIIFPamIRSUVH1&3	GAGTACCGGTTCTTTAATATGTTTACGGTCGTGATATG
IVFPamIRSUVH1&3	GAAACATATTAAGAACCGGACTCTACATATATATTCCT
IamirH13b	GATTAGGTAAAATAAGCATGCCCTCTCTCTTTTGTATTCC
IIamirH13b	GAGGGCATGCTTATTTTACCTAATCAAATAATCATGA
IIamirH13b	GAGGACATGCTTATTATACCTATTTACAGGTCGTGATATG
IVamirH13b	GAATAGGTATAATAAGCATGTCCTCTACATATATATTCCT
IImiRSUVH4	GATGAAAAAAAACGGTGTCCGACTCTCTCTTTTGTATTCC
IIImiRSUVH4	GAGTCGGACACCGTTTTTTTTTCATCAAAGAGAATCAATGA
IIImiRSUVH4	GAGCCGGACACCGTATTTTTTTCATCACAGGTCGTGATATG
IVmiRSUVH4	GATGAAAAAATACGGTGTCCGGC TCTACATATATATTCCT
IImiRSUVH4b	GATACCAGTTAAGTTATGCCGTCTCTCTCTTTTGTATTCC
IIImiRSUVH4b	GAGACGGCATAACTTAACTGGTATCAAAGAGAATCAATGA
IIImiRSUVH4b	GAGAAGGCATAACTTTACTGGTTTACAGGTCGTGATATG
IVmiRSUVH4b	GAAACCAGTAAAGTTATGCCTTCTCTACATATATATTCCT
IImiRSUVH5	GATTTAACTTTATACCGAGCGCTTCTCTCTTTTGTATTCC
IIImiRSUVH5	GAAGCGCTCGGTATAAAGTTAAATCAAAGAGAATCAATGA

IIIImiRSUVH5 GAAGAGCTCGGTATATAGTTAATTCACAGGTCGTGATATG  
 IVmiRSUVH5 GAATTA ACTATATACCGAGCTCTTCTACATATATATTCCT

IImiRSUVH6 GATTATAATCGATACCGCCTCGTCTCTCTTTTGTATTCC  
 IIImiRSUVH6 GACGAGGCCGGTATCGATTATAATCAAAGAGAATCAATGA  
 IIIImiRSUVH6 GACGCGGCCGGTATCCATTATATTCACAGGTCGTGATATG  
 IVmiRSUVH6 GAATATAATGGATACCGGCCGGTCTACATATATATTCCT

IFPamIRSUVH5&6 GATTATGGTTTATAAACCATGCCTTCTCTCTTTTGTATTCC  
 IIFPamIRSUV5&6 GAAGGCAGGTTTATAAACATAATCAAAGAGAATCAATGA  
 IIIFPamIRSUVH5&6 GAAGACAGGTTTATATAACCATATTCACAGGTCGTGATATG  
 IVFPamIRSUVH5&6 GAATATGGTATATAAACCTGTCTTCTACATATATATTCCT

lamirH56b GATTATGGTTTATAAACGTCCCTTCTCTCTTTTGTATTCC  
 IIamirH56b GAAGGGACGTTTATAAACATAATCAAAGAGAATCAATGA  
 IIIamirH56b GAAAGGACGTTTATTAACCATTATCACAGGTCGTGATATG  
 IVamirH56b GATAATGGTTAATAAACGTCCCTTCTACATATATATTCCT

IFPamIRSUVH7&8 GATATAAAAGTTCCGGCTCGTATTCTCTCTTTTGTATTCC  
 IIFPamIRSUV7&8 GAATACGAGCCGGAACTTTTATATCAAAGAGAATCAATGA  
 IIIFPamIRSUVH7&8 GAATCCGAGCCGGAAGTTTTATTTACAGGTCGTGATATG  
 IVFPamIRSUVH7&8 GAAATAAACTTCCGGCTCGGATTCTACATATATATTCCT

lamirH9 GATTAATACATACTCCGTTCCACTCTCTCTTTTGTATTCC  
 IIamirH9 GAGTGGAACGGAGTATGTATTAATCAAAGAGAATCAATGA  
 IIIamirH9 GAGTAGAACGGAGTAAGTATTATTCACAGGTCGTGATATG  
 IVamirH9 GAATAACTTACTACTCTACATATATATTCCT

IImiRSUVR1 GATTTTGTACGACTTGGGGCGCTTCTCTCTTTTGTATTCC  
 IIImiRSUVR1 GAAGCGCCCCAAGTCGTACAAAATCAAAGAGAATCAATGA  
 IIIImiRSUVR1 GAAGAGCCCCAAGTCCTACAAATTCACAGGTCGTGATATG  
 IVmiRSUVR1 GAATTTGTAGGACTTGGGGCTCTTCTACATATATATTCCT

IImiRSUVR2 GATATATGCAGATTAGGTGCCGTTCTCTCTTTTGTATTCC  
 IIImiRSUVR2 GAACGGCACCTAATCTGCATATATCAAAGAGAATCAATGA  
 IIIImiRSUVR2 GAACAGCACCTAATCAGCATATTTACAGGTCGTGATATG  
 IVmiRSUVR2 GAAATATGCTGATTAGGTGCTGTTCTACATATATATTCCT

IImiRSUVR4 GATTATATACGACCAATTGCCAGTCTCTCTTTTGTATTCC  
 IIImiRSUVR4 GACTGGCAATTGGTCGTATATAATCAAAGAGAATCAATGA  
 IIIImiRSUVR4 GACTAGCAATTGGTCCTATATATTCACAGGTCGTGATATG  
 IVmiRSUVR4 GAATATATAGGACCAATTGCTAGTCTACATATATATTCCT

IFPamIRSUVR1&2&4 GATTTTTCAGAACCCTAGTCTGCTCTCTCTTTTGTATTCC  
 IIFPamIRSUVR1&2&4 GAGCAGACTAGGGTTCTGAAAAATCAAAGAGAATCAATGA  
 IIIFPamIRSUVR1&2&4 GAGCCGACTAGGGTTGTGAAAAATTCACAGGTCGTGATATG  
 IVFPamIRSUVR1&2&4 GAATTTTACAACCCTAGTCGGCTCTACATATATATTCCT

IFPamIRSUVR5 GATTAAATTGAATTTCAAGCCGCTCTCTCTTTTGTATTCC  
 IIFPamIRSUVR5 GAGCGGGCTGAAATTCATTTAATCAAAGAGAATCAATGA

IIIFPamIRSUVR5 GAGCAGGCTGAAATTGAATTTATTCACAGGTCGTGATATG  
IVFPamIRSUVR5 GAATAAATTCAATTTACAGCCTGCTCTACATATATATTCCT

**Primers used for the RT-CAPS assay:**

ITS1for GCGCTACACTGATGTATTCAACGAG  
ITS1Rev CGCACCTTGCCTTCAAAGACTCGA

**Primers used for RT-PCR assays :**

pre-rRNA 3'ETS For CTCGAGGTTAAATGTTATTACTTGGTAAGATTCCGG  
pre-rRNA 3'ETS Rev TGGGTTTGTTCATATTGAACGTTTGTGTTTCATATCACC  
5Suvr4CDS For ATGATCAGTCTCTCCGACTAACC  
3Suvr4CDS Rev TCATTTGCGCTTTTTAGACACCTC  
5ACT2 for AAGTCATAACCATCGGAGCTG  
3ACT2 rev ACCAGATAAGACAAGACACAC

**Taqman primers for qPCR assays:**

ETStaqman For TGACATGGATTCTTCGAGGCCT  
ETStaqman Rev CATGACACGCCATTCTCTTCG  
A. arenosa ETS taqman probe Vic-TATATTGGGAAAGCGCATGGT-MBG  
A. thaliana ETS taqman probe 6Fam-TATATAACTTGTTTCGCATGATATT

Actin F: GAGAGATTGAGATGCCAGAAAGTC  
Actin R: TGGATTCCAGCAGCTTCCA  
AtSN1 F: CCAGAAATTCATCTTCTTTGGAAAAG  
AtSN1 R: GCCCAGTGGTAAATCTCTCAGATAGA  
5'ETS-4241F: TTGGAACGATTGATGATTTTGGAGT  
5'ETS-4316R: GCCTACGAACACTAGCTATCCGATC  
25S-10062F: TGTTCAACCCACCAATAGGGAA  
25S-10132R: TCAGTAGGGTAAAATAACCTGTCTCAC  
IGS-573F: GTCAAACACTTGGTGATATGAACACA  
IGS-623R: GCATGGGTTTGTTCATATTGAACGT  
3'ETS-84F: GAATTCCTCAACTTTACACGAGCTC  
3'ETS-134R: AAGTAATAACATTTAACCTCGAGAGACGAG

**A. thaliana-specific probe used for S1 nuclease protection**

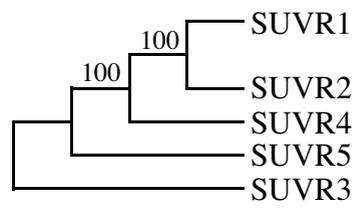
5'-GGGTTCCCCACGGACTGCCAGACTCCCTCAACACCCACCCCCCTATATAGCTGCC-3'

**PCR primers for generation of FISH probes**

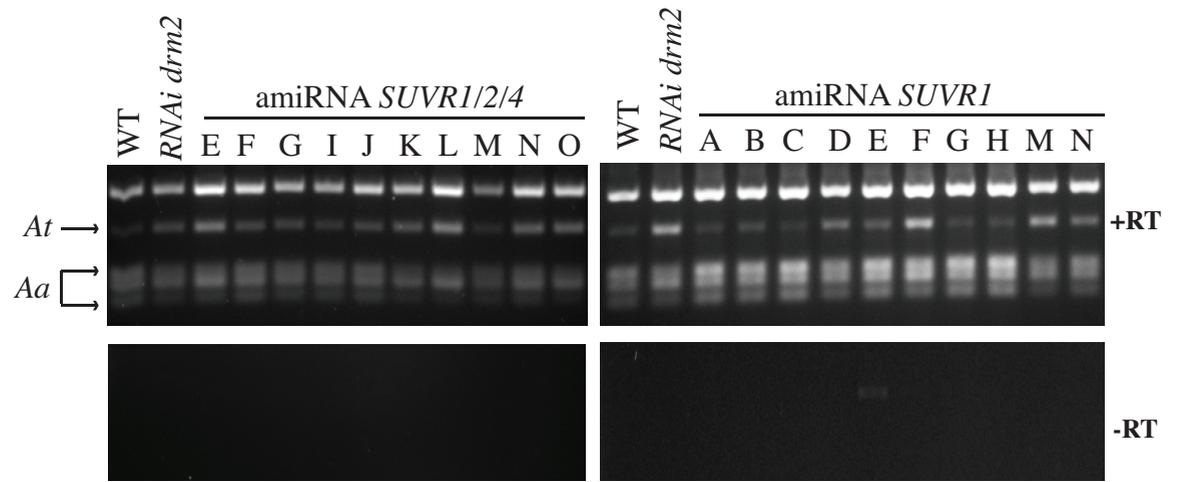
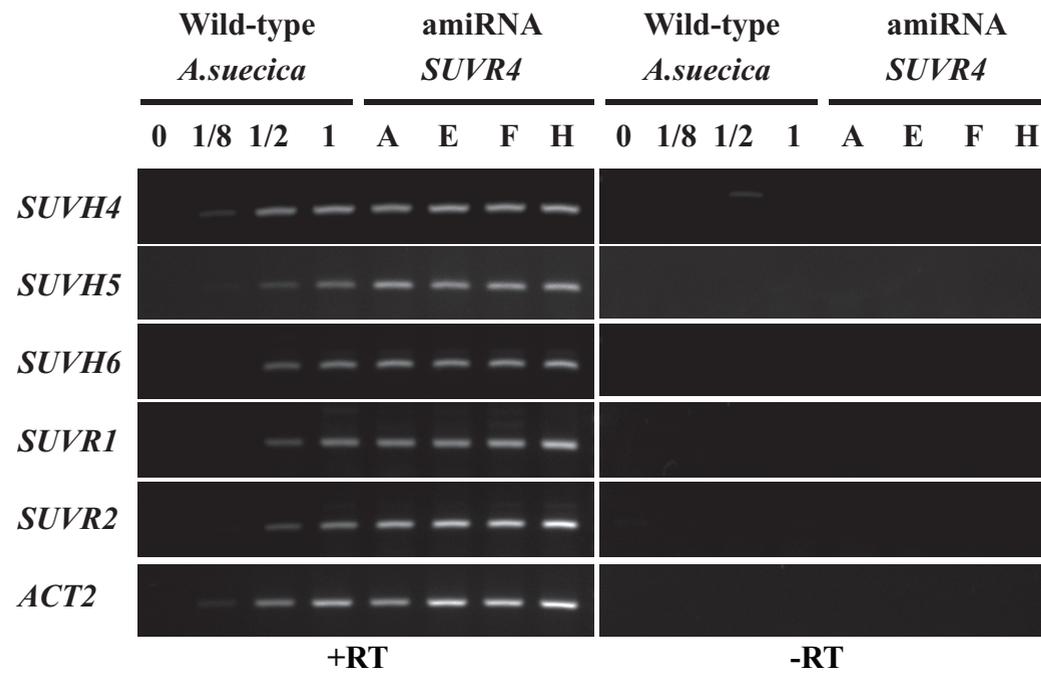
45S rRNA genes  
-250 to +250 +1 For CAAGCAAGCCCATTCTCCTC  
-250 to +250 +1 Rev CAACTAGACCATGAAAATCC

5S rRNA genes  
5Sfor GGATGCGATCATAACCAG  
5Srev CGAAAAGGTATCACATGCC

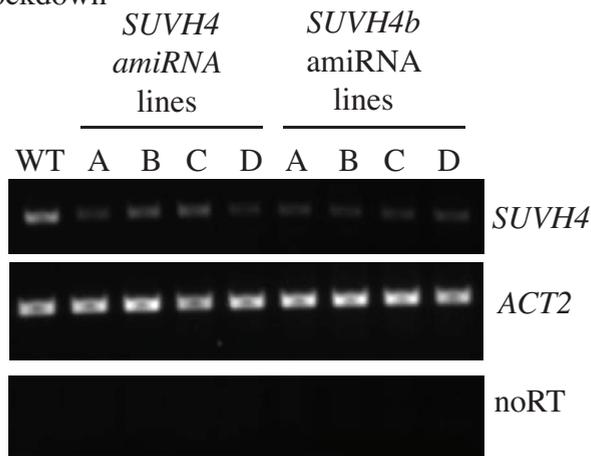
## A. Phylogenetic tree



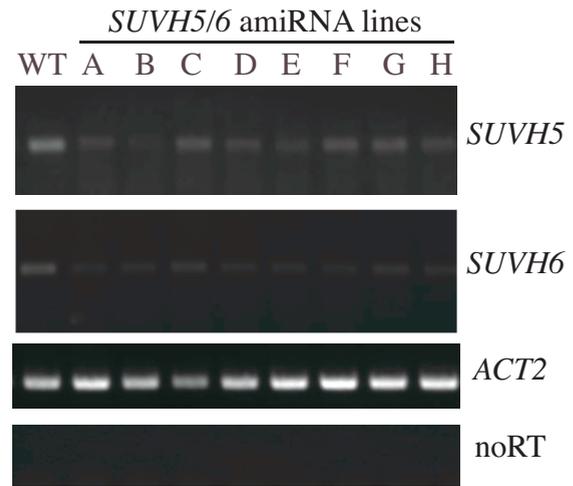
## B. RT-CAPS assay, T1 generation transgenic lines

C. RT-PCR amplification of *SUVR* or *SUVH* mRNAs in amiRNA lines targeting *SUVR4*.

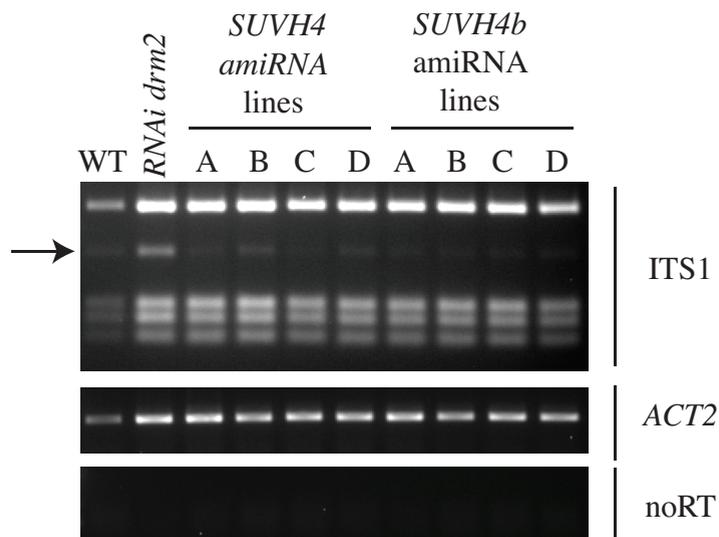
A. RT-PCR analysis of *SUVH4* mRNA knockdown



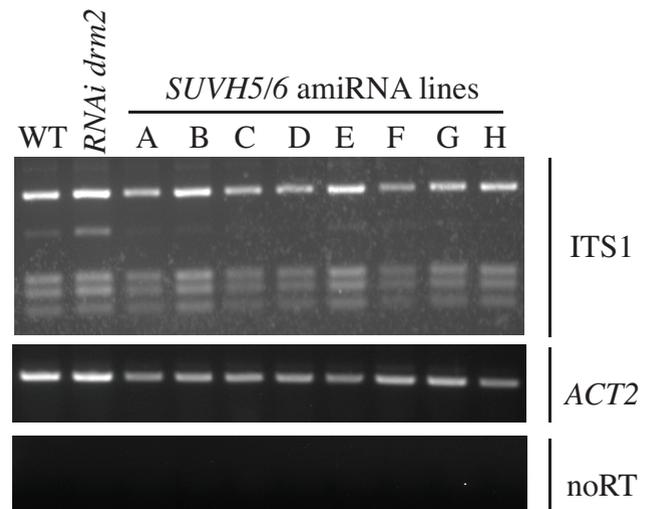
B. RT-PCR analysis of *SUVH5* and *SUVH6* mRNA knockdown



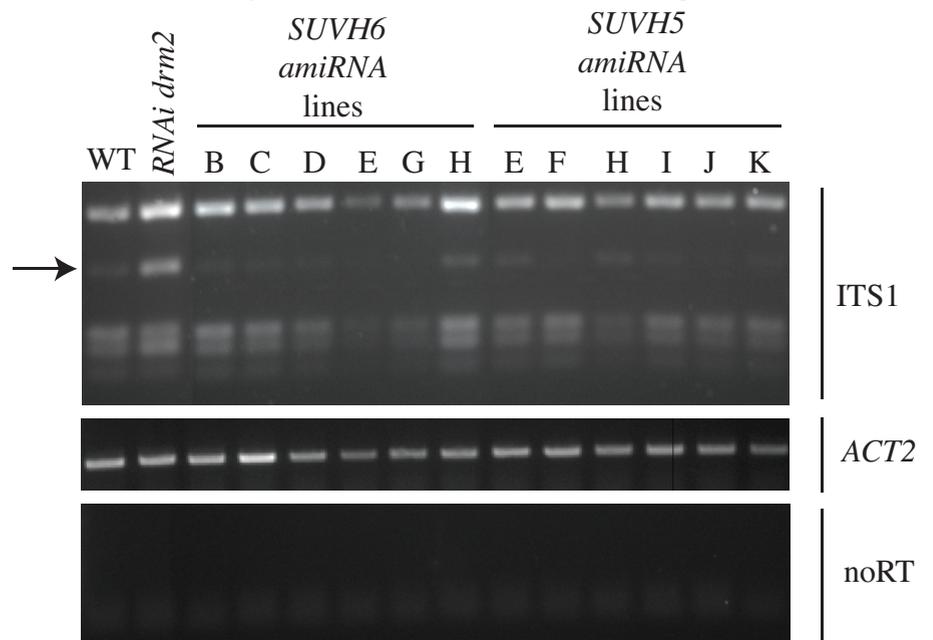
C. RT-CAPS analysis of nucleolar dominance



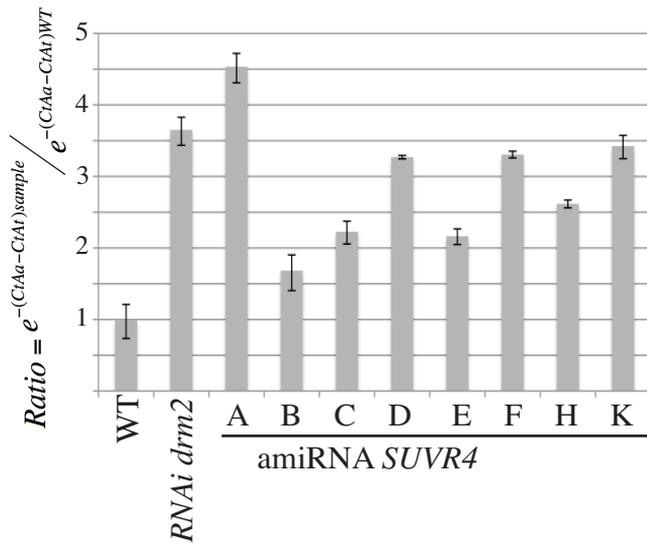
D. RT-CAPS analysis of nucleolar dominance; T1 generation



E. RT-CAPS analysis of nucleolar dominance; T1 generation



## qPCR analyses



## H3K27me1 and bulk H3 association with rRNA genes, determined by ChIP-seq

