# Science Advances <br> DAAAS 

## Supplementary Materials for

# ACD15, ACD21, and SLN regulate the accumulation and mobility of MBD6 to silence genes and transposable elements 

Brandon A. Boone et al.

Corresponding author: Steven E. Jacobsen, jacobsen@ucla.edu
Sci. Adv. 9, eadi9036 (2023)
DOI: 10.1126/sciadv.adi9036
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Figs. S1 to S6
Legend for table S1
Tables S2 and S3
Other Supplementary Material for this manuscript includes the following:
Table S1

A




B


|  | ACD15.5 |  | ACD21.4 |  | background |
| :---: | :---: | :---: | :---: | :---: | :---: |
| acd15.5 | guide $2+4$ | Insertion at guide 2 causing STOP codon | guide $2+4$ | WT | ColO |
| acd21.4 | guide $1+3$ | WT | guide $1+3$ | single base insertion (A) at guide 1 | ColO |
| acd15.5 acd21.4 | guide 1 + 3 | single base insertion (A) at guide 1 | guide $1+3$ | 37bp deletion at guide 1 causing STOP codon | ColO |
| adc15.5 acd21.4 sln | guide $1+3$ | single base deletion (A) at guide 1 | guide $1+3$ | $\qquad$ guide 1 | $\sin$ T-DNA: <br> SALK_090484 |


D


Fig. S1. ChIP-seq and RNA-seq analysis of ACD15 and ACD21.
A) Venn diagram of ChIP-seq peaks showing large overlap between samples. The peak sets indicated with circles (putative MBD5/6 unique, ACD15 unique, or MBD5/6/ACD15 unique peaks) were selected and visualized with heatmaps (right). We noted that at each peaks set groups, enrichment of most proteins was observed, thus suggesting that these regions are bound by all components of the MBD5/6 complex, despite not reaching our stringent significance threshold to be called as peaks. The heatmap shows $\log 2$ (fold-change) over no-FLAG control. B) Scheme of ACD15 and ACD21 genes showing the location of the guide RNAs used for CRISPR/Cas 9 mediated mutant generation. The table below shows the mutations obtained in each line. C) Bar plots showing the number of differentially expressed TEs (DE-TEs) or differentially expressed genes (DEGs) in the indicated genotypes. D) Upset plots showing the intersection of the upregulated genes or TEs found for each genotype. The largest intersection group constitutes loci upregulated in all six mutant lines.


Fig. S2. Organization of the MBD5/6 complex structure.
(A-I) Correlation between MBD6-RFP signal and either ACD15-YFP, ACD21-CFP, or SLNCFP signal in the indicated mutant backgrounds (underlined). Images represent individual zstack slices of roots from plants co-expressing MBD6 with either ACD15, ACD21, and SLN. Scatter plots indicate signal intensity for each fluorescent protein at each pixel of the image shown. Correlation coefficient: Pearson. Scale bars $=20 \mu \mathrm{M}$. (J-K) AlphaFold Multimer predicted structure of MBD5/6 complex with two copies each of MBD6, ACDC15, ACD21, and SLN along with confidence score map of the predicted complex. (L) Cartoon representation of the core dimeric MBD5/6 complex based on the AlphaFold Multimer prediction. The figure was created with Biorender.com.


Fig. S3. ACD15, ACD21, and SLN protein levels across plant lines.
(A) ACD15-YFP nuclear intensity in wild-type vs $\operatorname{sln}$ mutant plants. Comparisons were made using two-tailed t tests ( $\mathrm{NS}: \mathrm{P}>=0.05, \mathrm{~N}=52$ and 50 respectively). (B) Western blot analysis of ACD15-YFP protein levels along with ponceau stain as a loading control. (C) ACD21-CFP nuclear intensity in wild type vs $\operatorname{sln}$ mutant plants. Comparisons were made using two-tailed t tests (NS: $\mathrm{P}>=0.05, \mathrm{~N}=50$ per genotype). (D-E) Western blot analysis of ACD21-CFP protein levels along with ponceau stains as loading controls. (F) Western blot analysis of SLN-CFP protein levels along with ponceau staining as a loading control.

A
mbd5 mbd6


D

|  | FRAP Recovery Half Time ( $\mathrm{t}_{1 / 2}$ ) |  | Maximum Plateau Post Bleaching |  |
| :---: | :---: | :---: | :---: | :---: |
|  | Wild Type | sln | Wild Type | sln |
| MBD6 | $3.60(3.1755$ to 3.991$)$ | $2.44(1.813$ to 3.174$)$ | $0.93(0.9137$ ty 0.9561$)$ | $0.61(0.5826$ to 0.6336$)$ |
| ACD15 | $3.63(3.243$ to 4.046$)$ | $4.93(4.054$ to 5.951$)$ | $0.94(0.9233$ to 0.9635$)$ | $0.59(0.5676$ to 0.6212$)$ |
| ACD21 | $4.30(3.735$ to 4.927$)$ | $3.26(2.448$ to 4.220$)$ | $0.91(0.8813$ to 0.9351$)$ | $0.63(0.6009$ to 0.6625$)$ |

E



G



Fig. S4. SLN regulates the nuclear mobility of MBD5/6 complex members.
(A) Representative root nuclei images demonstrating MBD6-RFP overlap with ACD15-YFP, ACD21-CFP, and SLN-CFP. Scale bar $=2 \mu \mathrm{M}$. (B-C) MBD6-RFP signal within DAPI-stained nuclei. Scale bar $=2 \mu \mathrm{M}$. (D) Table of extrapolated values from FRAP curve data fitted with onephase association liner regression using GraphPad Prism. (E-H) FRAP curves of ACD15 and ACD21 along with representative nuclei images of FRAP experiments. Shaded area: 95\% confidence interval of FRAP data ( $\mathrm{N}=25$ from 5 plants lines), dots: mean values, line: fitted onephase, non-linear regression. Scale bars $=2 \mu$ M. (I-J) Intensity of ACD15 (I) and ACD21 (J) signal at 100 individual foci from multiple nuclei and plant lines. Comparisons were made using two-tailed t tests (****: $\mathrm{P}<0.0001$ ). (K) Expression of MBD6-RFP transgene across plant lines as measured by RT-qPCR. Comparisons were made using Welch's ANOVA and Dunnet's T3 multiple comparisons test $(* *: \mathrm{P}<0.01, \mathrm{NS}: \mathrm{P}>=0.05, \mathrm{~N}=4$ for wild type and 5 per mutant genotype). (L) MBD6-RFP nuclear intensity across wild type, acd15 acd21, and sln mutant plant lines. (NS: $\mathrm{P}>=0.05, \mathrm{~N}=50$ per genotype) (M) Representative nuclei showing MBD6-RFP foci in lil-1 mutant and control plants. Scale bars $=2 \mu \mathrm{M}$. (N) MBD6 foci counts across 50 slice Zstacks of root meristem from five plant lines per genotype. Two Tailed T-test (NS: $\mathrm{P}>=0.05$, $\mathrm{N}=5$ per genotype).

B

C
MBD6: MBD StkyC
MBD6 ${ }^{\text {NA }}$ MBD
MBD6 ${ }^{\text {CA }}=$ MBD

## G <br> 

D

E

H

I
mbd5 mbd6

L acd15 acd21


Fig. S5. The StkyC domain is required for MBD6's function.
(A) Protein structure representation of AlphaFold Multimer prediction of MBD6 with ACD15. Domains of MBD6 are annotated. (B) Protein alignment of MBD5, MBD6, and MBD7. MBDs and StkyC regions of MBD5, MBD6, and MBD7 are labelled. (C) Graphical representation of MBD6 deletion mutants. (D-E) RT-qPCR of $F W A$ expression comparing $m b d 5 m b d 6$ plants expressing MBD6 deletion mutants (D), along with representative images of MBD6 deletion mutants in root nuclei (E). Comparisons made using Brown-Forsythe ANOVA with Dunnett's multiple comparisons test ( ${ }^{* * *: ~} \mathrm{P}<0.001,{ }^{* *}: \mathrm{P}<0.01$, NS: $\mathrm{P}>=0.05, \mathrm{~N}=3$ per genotype. Scale bars $=2 \mu \mathrm{M})$. (F) RT-qPCR comparison of MBD6 deletion mutant expression compared to full length MBD6-RFP. Comparisons were made using Welch's ANOVA and Dunnet's T3 multiple comparisons test ( $* * *: ~ \mathrm{P}<0.001, \mathrm{NS}: \mathrm{P}>=0.05, \mathrm{~N}=4$ for wild type and 5 per MBD6 allele). (G) MBD6-RFP nuclear intensity of MBD6 deletion mutants in mbd5 mbd6 mutant plant lines. Comparisons made using Kruskal-Wallis test with Dunn's multiple comparisons test (NS: $\mathrm{P}>=0.05, \mathrm{~N}=50$ per MBD6 allele). (H-L) Correlation of MBD6-RFP domain, deletion mutants with ACD15-YFP or ACD21-CFP along with representative nuclear images. Correlation coefficient: Pearson. Scale bars $=2 \mu \mathrm{M}$.

A


B $\qquad$

C
Wild Type
SunTagTET1

D

$E$


G

HcFv-GFP-StkyC:

Fig. S6. SunTag ${ }^{\text {StkyC }}$ drives the formation of MBD5/6 nuclear foci.
(A-B) SunTag ${ }^{\text {StkyC }}$ expressing root nuclei stained with DAPI. Scale bars $=2 \mu \mathrm{M}$. (C)
Representative image of a root nucleus expressing SunTag ${ }^{\text {TET1 }}$. Scale bar $=2 \mu \mathrm{M}$. (D) Correlation of SunTag ${ }^{\text {StkyC }} F W A$ expression with leaf counts of individual T1 plants from Figure 5E-F. Correlation coefficient: Pearson. (E-G) RT-qPCR of $F W A$ in $m b d 5 m b d 6$ ( $\mathrm{N}=3,4$, and 7 respectively), $\operatorname{sln}(\mathrm{N}=3,4$, and 10 respectively), and $\operatorname{acd} 15 \operatorname{acd} 21$ ( $\mathrm{N}=3,3$, and 8 respectively) plants with and without SunTag ${ }^{\text {StkyC. Comparisons made using Brown-Forsythe ANOVA with }}$
 **: $\mathrm{P}<0.01$, *: $\mathrm{P}<0.05, \mathrm{NS}: \mathrm{P}>=0.05$ ). (H) Western blot analysis of scFv-GFP-StkyC using HA antibody along with ponceau staining as a loading control.

## Table S1. (Separate File)

Table showing merged IP-MS experiments. The numbers reported correspond to MS/MS counts. Col0 samples are no-FLAG negative controls. Each independent experiment is annotated with a shaded color.

| Protein | AtG ID |
| :---: | :---: |
| MBD6 | AT5G59380.1 |
| MBD5 | AT3G46580.1 |
| ACD15 | AT1G76440.1 |
| ACD21 | AT1G54850.1 |
| SLN | AT5G37380.1 |

Table S2. Gene identifiers for MBD5/6 complex.
All members of the MBD5/6 complex are listed along with gene identifiers associated with each gene for online $A$. thaliana database (https://www.arabidopsis.org/).

| Primer name | Sequence |
| :--- | :--- |
| FWA RT-qPCR Fw | TTAGATCCAAAGGAGTATCAAAG |
| FWA RT-qPCR Rev | CTTTGGTACCAGCGGAGA |
| IPP2 RT-qPCR Fw | GTATGAGTTGCTTCTCCAGCAAAG |
| IPP2 RT-qPCR Rev | GAGGATGGCTGCAACAAGTGT |
| RFP-qPCR Fw | ATGGCCTCCTCCGAGGACGTCATCA |
| RFP-qPCR Rev | CTTGGTCACCTTCAGCTTGGCGGTC |

Table S3. qPCR primer sequences.
List of qPCR primers, forward (Fw) and reverse (Rev), along with DNA sequences ( $5^{\prime}-3^{\prime}$ ).

